See reverse side for additional information

Interagency Report Control No 0180-DOA-AN

UNITED STATES DEPARTMENT OF AGRICULTURE ANIMAL AND PLANT HEALTH INSPECTION SERVICE

REGISTRATION NO. 23-R-0064

include Zip Code)

CUSTOMER NO. 353

2. HEADQUARTERS RESEARCH FACILITY (Name and Address, as registered with USDA,

FORM APPROVED OMB NO. 0579-0036

ANNUAL REPORT OF RESEARCH FACILITY

(TYPE OR PRINT)

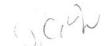
LEHIGH UNIVERSITY RESEARCH & SPONSORED PROGRAMS 526 BRODHEAD AVENUE #23B BETHLEHEM PA 18015

3. REPORTING FACILITY (List all locations where animals were housed or used in actual research, testing, teaching, or experimentation, or held for these purposes. Attach additional FACILITY LOCATIONS(sites) See Attached Listing 2006 (b)(2)High, (b)(7)f

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- 1) Professionally acceptable standards governing the care, treatment, and use of animals, including appropriate use of anesthetic, analgesic, and tranquilizing drugs, prior to, during, and following actual research, teaching, testing, surgery, or experimentation were followed by this research facility.
- 2) Each principal investigator has considered alternatives to painful procedures.
- 3) This facility is adhering to the standards and regulations under the Act, and it has required that exceptions to the standards and regulations be specified and explained by the principal investigator and approved by the Institutional Animal Care and Use Committee (IACUC). A summary of all the exceptions is attached to this annual report. In addition to identifying the IACUC-approved exceptions, this summary includes a brief explanation of the exceptions, as well as the species and number of animals affected.
- 4) The attending veterinarian for this research facility has appropriate authority to ensure the provision of adequate veterinary care and to oversee the adequacy of other

	CERTIFICATION BY HEADQUARTERS RESEARCH FACILITY OFFICIAL (Chief Executive Officer or Legally Responsible Institutional official)	-	
*			DATE SIGNED
Ā	(b)(6), (b)(7)c		11/16/06
			ADOLIABTEDE



Column E Explanation

This form is intended as an aid to completing the Column E explanation. It is not an official form and its use is voluntary. Names, addresses, protocols, veterinary care programs, and the like, are not required as part of an explanation. A Column E explanation must be written so as to be understood by lay persons as well as scientists.

	1. Registration Number: 23-R-0064	RECEIVED		
	2. Number 199of animals used in this study.	NOV 2 I 2006 BY:		
	3. Species (common name) Syrian Hamster of animals used in the study.			
•	 Explain the procedure producing pain and/or distress. Hamsters were food deprived between 6-48 hours without anest or analgesia. 	thetics		
	Please see attached "Exemption from Feeding Standard."			
5	Provide scientific justification why pain and/or distress could not be relieved. State determine that pain and/or distress relief would interfere with test results. (For Federal 1) For Federal 1.	methods or means used to derally mandated testing, see		
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ο.	What, if any, federal regulations require this procedure? Cite the agency, the code of Federal Regulations (CFR) title number and the specific section number (e.g., APHIS, 9 CFR 113.102):			
	AgencyCFR			

* EXEMPTION FROM FEEDING STANDARD 11/17/06*

In some experiments, animals are exempt from the feeding standard and analgesics are not administered. The exemption from feeding standard was approved because a body of research supported the contention that the 48 hours of food deprivation or food restriction in sheep did not cause pain and distress, and the animals maintained healthy equilibrium at a lower body fat content. Caloric restriction is used in our laboratory because we are studying the mechanisms that animals use to save energy during periods of low food availability to save energy by inhibition of the reproductive system. Nutritional infertility is an adaptive process whereby animals increase the ability to survive by resdistributing their energy economy. Metabolic fuels are shunted away from reproductive processes such as steroid production and mating, and are used for survival (Merry and Holehan, 1979; Merson and Kirkpatrick, 1981, Bronson, 1989). Animals have a sensory system designed to detect internal energy availability (Schneider, 1992). The sensory detectors of fuel availability send signals to parts of the brain that control fertility and ovulation as well as those areas that control hunger, food intake and caloric homeostasis. My experiments are designed to study the mechanisms that underlie this link between energy and fertility. The exemption from feeding standard allows hamsters and sheep to be fasted or food restricted to a degree that induces infertility and anestrus, but not to a degree that causes health problems.*

A 48-hour period of food deprivation has been commonly employed in research on rodents such as hamsters for many years (e.g., Roland et al., 1982; Schneider and Wade, 1989; Morin, 1986). For example, Dr. Morin (Morin, 1986) found that a 48-hour fast on days 1 and 2 of the estrous cycle induced anestrus, but this fast had no other health effects. Hamsters have a 4-day estrous cycle with ovulation and estrous behavior occurring on Day 4. One important piece of evidence demonstrating that a 48 h of fast is not harmful to Syrian hamsters is that when the 48 hour fast occurs on Days 2 and 3, Days 3 and 4, or on Days 4 and 1 or the cycle, there is no effect on estrous cyclicity (the hamsters showed normal ovulation rates and vigorous sex behavior). Ovulation and estrous behavior are inhibited only when the fast occurs on Days 1 and 2 of the estrous cycle, but not on any other days. Morin also found that normal estrous behavior could be easily reinstated in the 48 hour-fasted hamsters by giving an injection of estradiol. Furthermore, the effects on estrous cyclicity are rapidly reversed when the hamsters are returned to ad lib feeding. I subsequently demonstrated that the same 48-hour fast had no effect on estrous cyclicity or health in previously fattened hamsters (Schneider and Wade, 1989). Thus, a 48 hour fast has no effect on estrous cyclicity whatsoever when the hamsters have a high body fat content. In general, the fatter the hamster, the longer the period of food restriction necessary to induce anestrus. In sheep, much of the information comes from the work of my collaborator on adult ewes, e.g., Henry et al., 2001, 2004), or for peripubertal sheep from the laboratory of Technically, it does note violate the feeding standard because sheep eat one meal per day, and thus, this procedure does not require an exemption.

Analgesics are not appropriate because we are measuring indices of estrous cyclicity, which is under control of hypothalamic luteinizing hormone releasing hormone (LHRH), also known as gonadotropin releasing hormone (GnRH). LHRH and LH levels are adversely affected by opiates because it is well-known that opiates and opiate

derivatives, including analgesics such as buprenorphine, butorphenol and morphine, affect pituitary hormone secretion and hypothalamic LHRH (References 6-17 below). It is unreasonable to think that aspirin or nonsteroidal anti-inflammatory agents would be effective as analgesics on food deprived hamsters.

References

- 1. Schneider, J.E. and G.N. Wade. Availability of metabolic fuels controls estrous cyclicity of Syrian hamsters. *Science*, 244:1326-1328, 1989.
- 2. Morin, LP Environment and hamster reproduction: responses to phase-specific starvation during estrous cycle. Am. J. Physiol. 251:R663-R669, 1986.
- 3. Henry BA, Rao A, Tilbrook AJ, Clarke IJ. Chronic food-restriction alters the expression of somatostatin and growth hormone-releasing hormone in the ovariectomised ewe. J Endocrinol. 2001 Jul;170(1):R1-5.
- 4. Henry BA, Goding JW, Tilbrook AJ, Dunshea FR, Blache D, Clarke IJ. Leptin-mediated effects of undernutrition or fasting on luteinizing hormone and growth hormone secretion in ovariectomized ewes depend on the duration of metabolic perturbation. J Neuroendocrinol. 2004 Mar;16(3):244-55.
- 5. Rowland, N. Failure of deprived hamsters to increase food intake: some behavioral and physiological determinants. J. Comp. Physiol. Psychol. 96:591-603, 1982.
- 6. Ebling, F.J., Mirakhur, A., Maywood, .S. and Hastings, M.H. Photoperiodically induced changes in glutamatergic stimulation of LH secretion in male Syrian hamsters: role of circulating testosterone and endogenous opioids. Gen.Comp.Endocrinol. 96:50-62, 1994.
- 7. Mendelson, J.H., Ellingboe, J., Mello, N.K. and Kuehnle, J. Buprenorphine effects on plasma luteinizing hormone and prolactin in male heroin addicts. J.Pharmacol.Exp.Ther. 220:252-255, 1982.
- 8. Rolandi, E., Marabini, A., Franceschini, R., Messina, V., Bongera, P. and Barreca, T. Changes in pituitary secretion induced by an agonist-antagonist opioid drug buprenorphine. Acta. Endocrinol. 104:257-260, 1983.
- 9. Pechnick, R.N., George, R. and Poland, R.E. The effects of the acute administration of buprenorphine hydrochloride on the release of anterior pituitary hormones in the rat: evidence for the involvement of multiple opiate receptors. Life Sciences 37:1861-1868, 1985.
- 10. Pende, A., Musso, N.R., Montaldi, M.L., Pastorino, G., Arzese, M. and Deville, L. Evaluation of the effects induced by four opiate drugs, with different affinities to opioid receptor subtypes, on anterior pituitary LH, TSH, PRL and GH secretion and on cortisol secretion in normal men. Biomed.Pharmacother. 40:178-182, 1986.
- **11.** Fayez, M., Ahmed, H.H., el Nabarawy, F., Shokery, I.M. and el-Bauomy, A.M. Effects of butorphanol on luteinizing hormone and follicle stimulating hormone levels in ovariectomized rats---comparative study with morphine sulphate. Arch.Exp.Veterinarmed. 45:101-103, 1991.
- 12. Leshin, L.S., Rund, L.A., Kraeling, R.R. and Kiser, T.E. The bovine preoptic area and median eminence: sites of opioid inhibition of luteinizing hormone-releasing hormone secretion. J.Anim.Sci. 69:3733-3746, 1991.
- **13.** Okrasa, S., Kalamarz, H., Tilton, J.E. and Ziecik, A.J. Influence of opioids on LH secretion in gilts during the estrous cycle. J.Physiol.Pharmacol. 43:105-116, 1992.

- 14. Cicero, T.J., Nock, B. and O'Connor, L. Naloxone does not reverse the inhibitory effect of morphine on luteinizing hormone secretion in prepubescent male rats. J.Pharmacol.Exp.Ther. 264:47-53, 1993.
- 15. Adams, M.L., Sewing, B., Forman, J.B., Meyer, E.R. and Cicero, T.J. Opioid-induced suppression of at testicular function. J.Pharmacol.Exp.Ther. 266:323-328, 1993. 16. He, J.R., Molnar, J. and Barraclough, C.A. Morphine amplifies norepinephrine (NE)-induced LH release but blocks NE-stimulated increases in LHRH mRNA levels: comparison of responses obtained in ovariectomized, estrogen-treated normal and androgen-sterilized rats. Brain Res.Mol.Brain Res. 20:71-78, 1993.
- 17. Horton RJ, Li JY, Cummins JT, Smith AI, Shen PJ, Clarke IJ. Morphine decreases LH secretion in ovariectomized ewes only after steroid priming and not by direct pituitary action. Neuroendocrinology. 1990 Dec;52(6):612-7.

*In recent months in 2006, we have been investigating the general health of our breeding stock as well as a tendency for the hamsters to fail to show nutritional infertility, and so we have temporarily ceased using a 48 hour period of food deprivation in lean hamsters as an experimental group. At present time, our exemption from feeding standard will only apply to hamsters over 100 g in body weight until we increase the health of our hamsters colony.